



Total Synthesis of Bafilomycin A₁. 1. Syntheses of the C₅~C₁₁, C₁₂~C₁₇ and C₁₈~C₂₅ Segments.

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Abstract: The effective syntheses of the C₅~C₁₁ (2), C₁₂~C₁₇ (3) and C₁₈~C₂₅ (4) segments, which are promising synthetic intermediates toward the total synthesis of the macrolide antibiotic, bafilomycin A₁ (1), were described.

Bafilomycin A₁ (**1**) isolated in 1983 by Werner and Hagenmaier¹ is the first specific potent inhibitor of vacuolar H⁺-ATPase.² The structure and absolute configuration of **1** were established by X-ray crystallographic analysis³ and by NMR spectroscopy.⁴ Bafilomycin A₁ (**1**) belongs to a family of structurally related polyketide macrolide antibiotics. The other macrolide antibiotics such as elaiophylin,⁵ the concanamycins⁶ and the hygrolidins⁷ are closely related to the bafilomycins. The most unique and striking structural feature of these macrolide families is an unusual 16- or 18-membered tetraenic lactone ring with a β-hydroxy hemiacetal side chain. An efficient aldol method for the assembly of **1** was recently reported by Evans and Calter,⁸ and elegant syntheses of the C₁₃~C₂₅ segments of **1** have been independently announced by Roush's⁹ and Paterson's¹⁰ groups. Herein we disclose the effective syntheses of the C₅~C₁₁ (**2**), C₁₂~C₁₇ (**3**) and C₁₈~C₂₅ (**4**) segments which are promising synthetic intermediates toward the total synthesis¹¹ of the biologically important natural product, bafilomycin A₁ (**1**) (Figure 1).

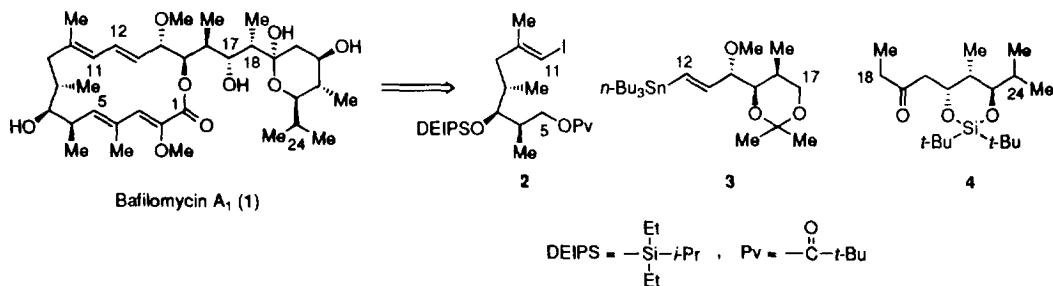
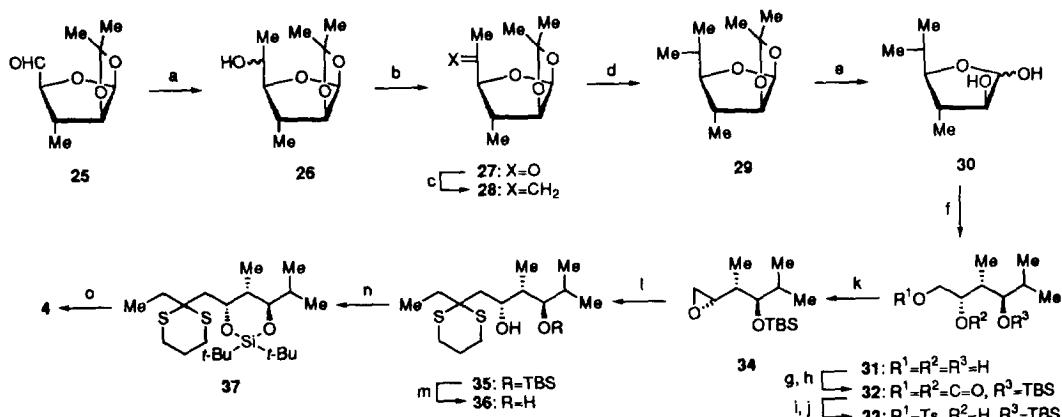
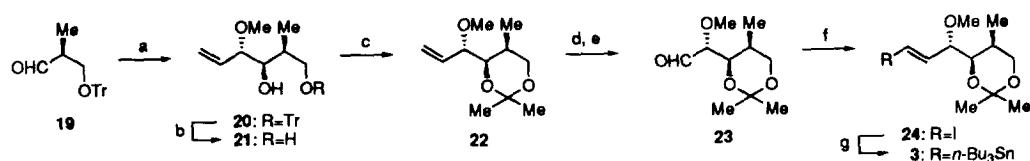
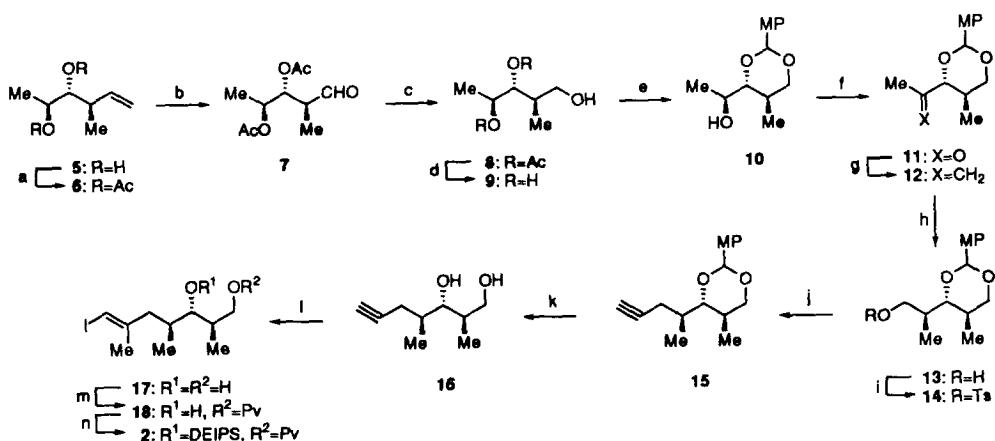


Figure 1

Synthesis of the C5~C11 segment 2. The synthesis of the vinyl iodide **2** corresponding to the C5~C11 segment of baflomycin A1 (**1**) is summarized in Scheme 1. The starting material **5**¹² was first converted into the triol **9** via oxidative cleavage of the double bond in **6** in four steps (1. Ac₂O, 4-DMAP, EtOAc, r. t., 1 h; 2. O₃, MeOH-CH₂Cl₂, -78 °C, 2 h then Me₂S; 3. NaBH₄, MeOH-CH₂Cl₂, r. t., 0.5 h; 4. NaOMe, MeOH, r. t., 3 h) in 94% overall yield. Selective *p*-methoxybenzylideneation of the 1,3-diol in **9** (*p*-methoxybenzaldehyde dimethyl acetal, CSA, DMF, r. t., 1.5 h), followed by oxidation (PCC, MS 3A, CH₂Cl₂, r. t., 1.5 h) and the Wittig reaction using Ph₃P=CH₂ in benzene afforded **12** in 89% overall yield. Hydroboration of **12** employing dicyclohexylborane in THF at room temperature for 1 h proceeded with complete stereoselectivity to give only the alcohol **13** in 88% yield after the subsequent oxidative workup. Tosylation of the alcohol **13** (TsCl, Py, r. t., 1.5 h) yielded the tosylate **14** which was subjected to the reaction with lithium acetylide (5 equiv.) in dimethyl sulfoxide to give the acetylene **15** in 66% overall yield. After deprotection of the *p*-methoxybenzylidene group in **15** under acidic conditions (80% AcOH-H₂O, 40 °C, 13 h), the resultant diol **16** was treated with Cp₂ZrCl₂, Me₃Al and I₂ in 1,2-dichloroethane¹³ to afford only the tri-substituted *trans* vinyl iodide **17** in 63% overall yield from **15**. Selective pivaloylation (PvCl, Et₃N, CH₂Cl₂, r. t., 14 h) of the primary alcohol in **17**, followed by silylation (DEIPSOTf, 2,6-lutidine, CH₂Cl₂, r. t., 4 h) with the diethylisopropylsilyl (DEIPS) group¹⁴ furnished the suitably protected vinyl iodide **2** in 97% overall yield.

Synthesis of the C12~C17 segment 3. Scheme 2 illustrates the synthesis of the vinyl butyltin **3** corresponding to the C12~C17 segment of baflomycin A1 (**1**). Treatment of the aldehyde **19**¹⁵ with *in situ* generated γ -methoxyallylchromium reagent^{9,16} (CrCl₂, CH₂=CHCH(OMe)₂, TMS-I, THF, -42 °C, 16 h) afforded the homoallylic alcohol **20** as a major diastereomer with 10 : 1.1 : 0.5 selectivity in 62% yield. The trityl group in **20** was removed under acidic conditions (1% HCl-MeOH, r. t., 0.5 h) and the resultant diol **21** was then protected with an isopropylidene group (2,2-dimethoxypropane, CSA, CH₂Cl₂, r. t., 16 h) to provide the acetonide **22**. Dihydroxylation (OsO₄, NMO, acetone-H₂O, r. t., 16 h) of **22**, followed by sequential periodate-oxidation (NaIO₄, THF-H₂O, r. t., 0.5 h) and Takai's reaction¹⁷ using CrCl₂ and CHI₃ in THF gave only the *trans* vinyl iodide **24** in 38% overall yield from **20**. Finally, treatment of **24** with *n*-Bu₃SnCl and *n*-BuLi in THF at -78 °C for 1 h afforded the vinyl butyltin **3** in 69% yield.

Synthesis of the C18~C25 segment 4. The synthesis of the ethyl ketone **4** corresponding to the C18~C25 segment of baflomycin A1 (**1**) is depicted in Scheme 3. This synthesis began with the conversion of the sugar derivative **25**¹⁸ into **29** possessing an isopropyl group in standard manners in four steps (1. MeMgI, Et₂O, r. t., 0.5 h; 2. PCC, MS 3A, CH₂Cl₂, r. t., 0.5 h; 3. Ph₃P=CH₂, benzene, r. t., 0.5 h; 4. H₂, Raney-Ni(W4), dioxane, r. t., 24 h) and in 66% overall yield. Hydrolysis (50% AcOH-H₂O, 80 °C, 2 h) of the 1,2-isopropylidene group in **29**, followed by lithiumaluminum hydride (LAH)-reduction afforded the triol **31** which was then subjected to the selective protection of the 1,2-diol with carbonate (*N,N'*-carbonyldiimidazole, CH₂Cl₂, r. t., 2.5 h) and silylation (TBS-Cl, imid., DMF, 40 °C, 16 h) of the resultant secondary alcohol to provide **32** in 52% overall yield from **29**. After removal of the carbonate group in **32** by hydrolysis (1*N* NaOH, MeOH, r. t., 16 h), selective tosylation (TsCl, Py, r. t., 16 h) of the resultant primary alcohol and sequential epoxidation (NaOMe, MeOH-CHCl₃, r. t., 16 h) gave the epoxide **34** in 46% overall yield. Reaction of **34** with 2-ethyl-2-lithio-1,3-dithiane (5 equiv.) in THF at -20 °C for 1 h afforded the dithioacetal **35** whose silyl group was removed (TBAF, THF, r. t., 2.5 h) to give the diol **36** in 97% overall yield. Finally, the diol **36** was protected (*t*-Bu₂Si(OTf)₂, DMF, r. t., 2 h) with a di-*t*-butylsilyl group and then the dithioacetal



group was cleaved (CaCO₃, MeI, MeCN-H₂O, r. t., 6 h) to furnish the ethyl ketone **4⁸** in 66% overall yield.¹⁹

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- All new compounds were purified by silica-gel column chromatography and were fully characterized by spectroscopic means. Selected ¹H-NMR spectra [270MHz, CDCl₃, δ (TMS), J (Hz)] are the following. **2**: 0.64 (4H, q, *J* = 7.7), 0.86 (3H, d, *J* = 7.0), 0.95-1.03 (16H, m), 1.21 (9H, s), 1.80 (3H, s), 1.8-1.93 (1H, m), 1.95-2.07 (1H, m), 2.02 (1H, dd, *J* = 13.0 and 10.8), 2.42 (1H, br dd, *J* = 13.0 and 3.0), 3.51 (1H, dd, *J* = 4.5 and 4.5), 3.87 (1H, dd, *J* = 11.0 and 7.3), 4.25 (1H, dd, *J* = 11.0 and 4.9), 5.85 (1H, br s). **3**: 0.85-0.93 (15H, m), 1.10 (3H, d, *J* = 7.0), 1.24-1.38 (6H, m), 1.35 (3H, s), 1.37 (3H, s), 1.44-1.57 (6H, m), 1.77 (1H, m), 3.28 (3H, s), 3.40 (1H, ddd, *J* = 9.0, 6.3 and 1.2), 3.60 (1H, dd, *J* = 11.6 and 1.8), 3.82 (1H, dd, *J* = 9.0 and 2.2), 4.08 (1H, dd, *J* = 11.6 and 2.8), 5.79 (1H, dd, *J* = 19.0 and 6.3), 6.21 (1H, dd, *J* = 19.0 and 1.2). **4**: 0.73 (3H, d, *J* = 7.0), 0.86 (3H, d, *J* = 6.9), 0.97 (9H, s), 0.99 (9H, s), 1.01 (3H, d, *J* = 7.0), 1.07 (3H, d, *J* = 7.1), 1.73 (1H, d septet, *J* = 7.0 and 2.1), 2.23 (1H, m), 2.38 (1H, dd, *J* = 14.2 and 3.4), 2.53 (1H, dq, *J* = 10.8 and 7.0), 2.56 (1H, dq, *J* = 10.8 and 7.0), 2.71 (1H, dd, *J* = 14.2 and 10.1), 3.68 (1H, dd, *J* = 9.6 and 2.1), 4.62 (1H, ddd, *J* = 10.1, 5.9 and 3.4).

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